

Hematopoietic stem cell stretches and moves in its bone marrow niche

Walison N. Silva^{a,1}, Alinne C. Costa^{a,1}, Caroline C. Picoli^{a,1}, Beatriz G.S. Rocha^a, Gabryella S. P. Santos^a, Pedro A.C. Costa^a, Parviz Azimnasab-sorkhabi^a, Maryam Soltani-asl^a, Rodrigo A. da Silva^b, Jaime Henrique Amorim^c, Rodrigo R. Resende^d, Akiva Mintz^e, Alexander Birbrair^{a,e,*}

^a Department of Pathology, Federal University of Minas Gerais, Belo Horizonte, MG, Brazil

^b Department of Dentistry, University of Taubaté, Taubaté, São Paulo, Brazil

^c Center of Biological Sciences and Health, Federal University of West Bahia, Barreiras, BA, Brazil

^d Department of Biochemistry and Immunology, Federal University of Minas Gerais, Belo Horizonte, MG, Brazil

^e Department of Radiology, Columbia University Medical Center, New York, NY, USA

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ABSTRACT

Hematopoietic stem cells are the most illustrious inhabitants of the bone marrow. Direct visualization of endogenous hematopoietic stem cells in this niche is essential to study their functions. Until recently this was not possible in live animals. Recent studies, using state-of-the-art technologies, including sophisticated in vivo inducible genetic approaches in combination with two-photon laser scanning microscopy, allow the follow-up of endogenous hematopoietic stem cells' behavior in their habitat. Strikingly, the new findings reveal that quiescent hematopoietic stem cells are more mobile than previously thought, and link their retained steady state within the niche to a mobile behavior. The arising knowledge from this research will be critical for the therapy of several hematological diseases. Here, we review recent progress in our understanding of hematopoietic stem cell biology in their niches.

1. Introduction

1.1. Hematopoietic stem cells

The bone marrow is, presently, well-established as the primary postnatal site of new blood cells formation, generating approximately 10 billion leukocytes, 200 billion red cells, and 400 billion platelets daily during our whole life (Yoder, 2004). Nevertheless, this bone marrow's capacity was only first experimentally discovered at the second half of the 19th century by a German pathologist Ernst Neumann (Cooper, 2011). He also proposed the controversial, at that time, concept that one cell type may originate all other blood cells in the bone marrow (Cooper, 2011). This pioneer theory introduced the field of hematopoietic stem cell biology. Early works from the 50 s demonstrated that transplantation of bone marrow cells could protect the organism from some of the damages caused by irradiation, avoiding hematopoietic failure (Jacobson et al., 1950; Lorenz et al., 1951; Jacobson et al., 1951; Main and Prehn, 1955), suggesting the existence of a cell with reconstitutive ability in the middle of bone marrow cells. In the 60 s, James Till, Ernest

McCulloch, and their colleagues brought the initial experimental proof of the existence of hematopoietic stem cells. They demonstrated that there were cells in the bone marrow with capacity to generate all blood cell types and make more of themselves (Till and Mc, 1961; Wu et al., 1968; Becker et al., 1963; Siminovitch et al., 1963; Wu et al., 1967; Wolf and Trentin, 1970). Since then, in the clinic, intravenous transplantation of bone marrow cells has proven to be effective to treat patients with several blood-related diseases, such as leukemia (Thomas et al., 1957; Mathe et al., 1959, 1963). In leukemic patients, bone marrow transplantation has revolutionized therapeutic options, and now is widely used in the clinic, allowing bone marrow cells from healthy donors to repopulate the bones of patients with leukemia after aggressive chemotherapy (Wayne et al., 2010; Yamamoto et al., 2020; Khaddour et al., 2020; Sanchez-Aguilera and Mendez-Ferrer, 2017; Singh and Cancelas, 2020).

Nowadays, hematopoietic stem cells can be isolated from the bone marrow highly enriched by using multiple specific molecular markers (Laurenti et al., 2008). Scientists are constantly searching for new markers to isolate subsets of purified hematopoietic stem cells. It is well

* Corresponding author at: Department of Pathology, Federal University of Minas Gerais, Belo Horizonte, Brazil.

¹ Co-first authors.

accepted that stem cells capable of hematopoietic reconstitution are positive for Sca-1, a membrane glycoprotein (Okada et al., 1992) and c-Kit, a tyrosine kinase receptor (CD117), concomitantly being negative for lineage markers (Lin⁻), including Gr-1, Ter119, Mac-1, B220, CD4 and CD8 (Challen et al., 2009; Kiel et al., 2007a; Dykstra et al., 2006). Additionally, these characteristics are combined with strategies established by different groups to isolate purified hematopoietic stem cells (Kiel et al., 2007a, b), such as their status of expression of Thy1.1, Flk2, CD34, Endoglin (CD105) (Chen et al., 2002), Tie-2 (Arai et al., 2004), endothelial protein C receptor (EPCR) (Balazs et al., 2006), CD244, CD48, and/or CD150 (Kiel et al., 2007a). The exclusion of fluorescent dyes is an additional method that has proven advantageous to select for cells enriched with hematopoietic stem cells activity (Okada et al., 1992; Challen et al., 2009; Goodell et al., 1997; Pearce et al., 2004).

One obstacle in the hematopoietic stem cells' isolation is that the number of available compatible bone marrow donors still limits the usage of hematopoietic stem cells for transplantation. Although hematopoietic stem cells are maintained throughout all our life in their niche *in vivo*, we still are unable to multiply and expand effectively hematopoietic stem cells *in vitro* under suitable conditions. Therefore, a deeper understanding of hematopoietic stem cells biology will be essential for the better efficiency of bone marrow transplantation in the future.

In this review, we discuss the recent progress in our understanding of hematopoietic stem cell biology in their niches, focusing on hematopoietic stem cells' heterogeneity and interactions with other cells in the context of recent findings. Furthermore, we shed light on the gaps in the field and highlight important open questions.

1.2. Hematopoietic stem cells within the bone marrow niche

Hematopoietic stem cells reside predominantly within the bone marrow (Birbrair and Frenette, 2016a). The hematopoietic stem cells' bone marrow niche regulates the behavior of those cells (Schofield, 1978). Hematopoietic stem cell fate is decided by the pro-quiescence, pro-renewal, or pro-differentiation intrinsic and extrinsic regulators inside the niche (Rashidi et al., 2014). Multiple genetically engineered mouse models have been extensively used to explore the complexity of the hematopoietic stem cell niche within the bone marrow. These investigations established diverse components as niche-supporting cells for hematopoietic stem cells, providing many molecules, such as cytokines, to control hematopoietic stem cell function (Asada et al., 2017a). Experimental proof has revealed that intervention in the key niche regulators may lead to various hematologic pathologic processes (Birbrair and Frenette, 2016a; Borges et al., 2017). Thus, understanding hematopoietic stem cells' behavior in their niche, as well as their interactions with other niche constituents, is of crucial significance.

Direct visualization of hematopoietic stem cells in their niche is necessary to study their activity *in vivo*. This was possible with the advancement of deep confocal microscopic imaging that helped determine hematopoietic stem cell niche architecture. Several studies analyzed the localization of hematopoietic stem cells relative to distinct niche components (Kunisaki et al., 2013; Asada et al., 2017b; Acar et al., 2015). In most studies, the hematopoietic stem cells behavior was analyzed in bone marrow biopsies, in which hematopoietic stem cells can be precisely identified using a combination of molecular markers by immunohistochemistry (Kunisaki et al., 2013; Asada et al., 2017b; Acar et al., 2015). Nevertheless, remains the open question whether hematopoietic stem cell behavior is the same within the bones of live animals. Other works analyzed the behavior of pre-labeled hematopoietic stem cells in recipient live mice (Lo Celso et al., 2009; Lewandowski et al., 2010; Takizawa et al., 2011). Nevertheless, it is not clear whether the non-physiological behavior of these introduced hematopoietic stem cells is the same as of endogenous stem cells. Additionally, for the efficiency of transplantation, recipient animals receive treatments that affect the bone marrow microenvironment, bringing the possibility of

changes in hematopoietic stem cell behavior due to niche disruption.

Now, in a recent article in *Cell Stem Cell*, Upadhaya and colleagues demonstrated elegantly how endogenous adult hematopoietic stem cells behave in the bone marrow in live animals (Upadhaya et al., 2020). Using state-of-the-art technologies, including sophisticated *in vivo* inducible genetic approaches, such as lineage-tracing Cre/loxP mediated technologies, in combination with two-photon laser scanning microscopy, the authors selectively followed the behavior of single adult hematopoietic stem cells for several hours. The authors analyzed the bone marrow of a mouse model in which specifically endogenous hematopoietic stem cells produce red fluorescence, Pdzk1ip1-CreER/TdTomato mice. Behaviors of hematopoietic stem cells and macrophages, which were detected by their autofluorescence, were compared. These experiments revealed that hematopoietic stem cells present a constantly changing not-rounded shape extending cytoplasmic projections, in contrast to the perfectly round cells as previously thought. Surprisingly, hematopoietic stem cells moved 7.5 times more than resident macrophages in steady state conditions (Upadhaya et al., 2020) (Fig. 1). Importantly, the authors confirmed that Pdzk1ip1-expressing cells were *bona fide* hematopoietic stem cells by confirming that the investigated cells were also Fgd5⁺ in Pdzk1ip1-CreER/TdTomato/Fgd5-ZsGreen mice. Upadhaya and colleagues also reported, as previously known, that hematopoietic stem cells are located in the perivascular space, and physically interact with stem cell factor (SCF)-expressing pericytes in the bone marrow. Strikingly, mobilization of the hematopoietic stem cells from the bone marrow niche by drugs that block C-X-C chemokine receptor type 4 (CXCR4) receptor and integrin signaling inhibited hematopoietic stem cell mobility as well as its form fluctuations within the niche (Upadhaya et al., 2020) (Fig. 2). This study reveals that hematopoietic stem cells are more mobile than previously thought, and links their retained steady state within the niche to a mobile behavior. Here, we discuss the findings from this work and evaluate recent advances in our understanding of the hematopoietic stem cell microenvironment.

2. Perspectives / future directions

2.1. Hematopoietic stem cells heterogeneity

Hematopoietic stem cells are not homogeneous. There have been shown subpopulations based on their life span (Yang et al., 2005), specific surface markers (Birbrair and Frenette, 2016b), differentiation capacities (Muller-Sieburg et al., 2012), and level of self-renewal (Ema et al., 2005). Although great advances were made regarding our knowledge of the bone marrow niche components, how extrinsic regulators act on hematopoietic stem cell subsets remains completely unknown. Interestingly, Upadhaya and colleagues analyzed only about one-fifth of hematopoietic stem cells, as this is approximately the amount labeled in Pdzk1ip1-CreER/TdTomato mice (Upadhaya et al., 2020). It remains unclear whether in these transgenic mice a subpopulation of rapidly moving hematopoietic stem cells is selected or whether all hematopoietic stem cells display approximately the same rate of movement. Future studies should study the behavior of not-expressing Pdzk1ip1 hematopoietic stem cells.

Hematopoietic stem cells modify their differentiation capacity during aging, losing gradually their self-renewal ability, becoming increasingly myeloid-biased (Mendelson and Frenette, 2014; Pang et al., 2011). The changes perceived in old hematopoietic stem cells were speculated to be exclusively due to hematopoietic stem cell-intrinsic alterations (Geiger et al., 2013; Birbrair et al., 2013). Nonetheless, recent results show the critical function of several extrinsic molecules inducing hematopoietic stem cell aging as well (Nakamura-Ishizu and Suda, 2014). It will be interesting to explore how hematopoietic stem cells' behavior changes in live animals with aging, and whether myeloid-biased hematopoietic stem cells behave differently from the others.

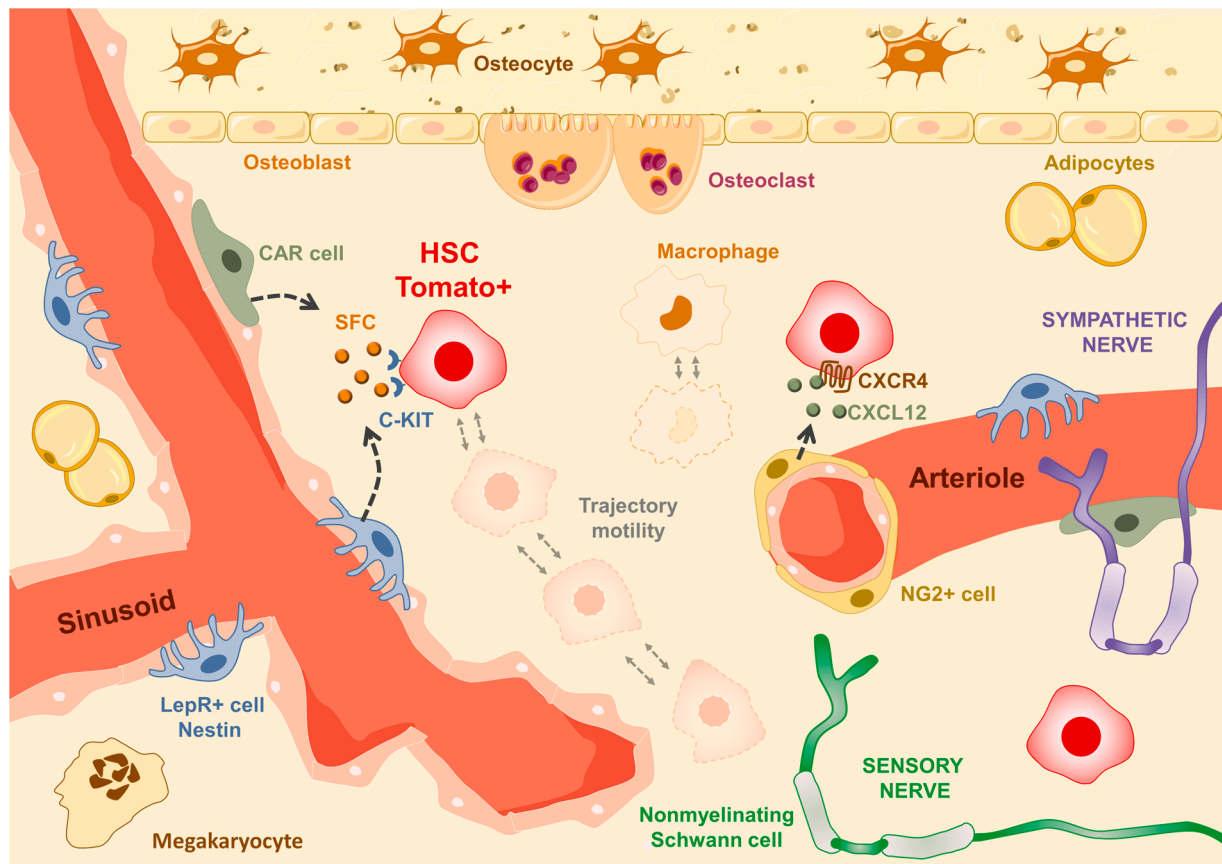


Fig. 1. Schematic illustrating hematopoietic stem cell movement within the bone marrow niche.

Hematopoietic stem cells (in red) present dynamic morphology (non-spherical) and complex motile behavior when compared to sessile resident macrophages (in brown) within the bone marrow cavity. Upadhaya and colleagues demonstrated that hematopoietic stem cells' displacement velocity is 7.5 times faster than macrophages (Upadhaya et al., 2020).

2.2. Other hematopoietic stem cell niches

During embryonic development, hematopoiesis occurs at specific anatomical sites that change with the developmental age (Khan et al., 2016; Al-Drees et al., 2015; Palis et al., 1999; Tavian and Peault, 2005). This happens because of the migration of hematopoietic stem cells throughout the embryo (Bowman and Zon, 2009). The hematopoietic activity starts in the extraembryonic yolk sac at embryonic day 7.5; then, at day 9, it advances to the dorsal aorta-gonad-mesonephros (AGM region), the para-aortic splanchnopleura, and chorioallantoic placenta (Rhodes et al., 2008); at day 10, it arrives to vitelline and umbilical arteries, spleen, skeletal muscle surrounding the developing long bones, and the fetal liver, where hematopoietic stem cells expand exponentially (Al-Drees et al., 2015; Palis et al., 1999; Swain et al., 2014; Tanaka et al., 2015; Medvinsky et al., 1993, 2011; Baron, 2005; Baron et al., 2012; Barminko et al., 2016; Kumaravelu et al., 2002; Muller et al., 1994; Medvinsky and Dzierzak, 1996; Sugiyama and Tsuji, 2006; Lux et al., 2008). Lastly, at day 15, hematopoietic stem cells from the fetal liver move through the circulation to the bone marrow cavity, which turns into the dominant niche for hematopoietic stem cells throughout the whole adult life (Khan et al., 2016; Medvinsky et al., 2011). Hematopoietic stem cells in adults can also appear outside the medullary spaces. This phenomenon is termed extramedullary hematopoiesis, was reported in adults in the periosteum, spleen, liver, heart, kidney, adrenal glands, fatty tissue, intra-spinal tissue, para-vertebral regions, pre-sacral region, nasopharyngeal region, paranasal sinuses, and in multiple types of cancers (Sohawon et al., 2012; Johns and Christopher, 2012; Tsamandas et al., 1995; Vassiliou et al., 2012; Macki et al., 2013; Inra et al., 2015; Bozzini et al., 1970; Bowen et al., 2015; Schnuelle et al., 1999;

Woodward et al., 2000; Lewis et al., 1994). Although it normally indicates a pathologic state of the organ, recent works show the extramedullary hematopoiesis may occur under physiologic conditions as well. Elegant studies have shown the presence of hematopoietic stem cells in the pulmonary microenvironment under physiologic circumstances (Borges et al., 2017; Lefrancais et al., 2017). Future studies using modern technologies such as two-photon laser scanning microscopy adapted to the specific organs will reveal how hematopoietic stem cells behave in these extramedullary niches.

2.3. The quiescent state

The definition of quiescence emerged from the perception that each cell in a population proliferates at its own rate (Cheung and Rando, 2013). Thus, cells that are in a non-proliferative state are termed quiescent, even under certain stimuli they can enter the cell cycle and start proliferating. Unicellular organisms, which survive in adverse habitats, enter the quiescent state to not be extinct (Gray et al., 2004). Similarly, stem cells exist in a quiescent state throughout our life to keep for as long as possible a reserve pool. Despite quiescence being considered as a dormant static state, quiescence seems to portray a state in which the stem cell is ready to be activated. Upadhaya and colleagues demonstrate that quiescent hematopoietic stem cells are not so "dormant", being rather "awake" based on the movement that they present within the niche (Upadhaya et al., 2020). The reason for this augmented mobility of hematopoietic stem cells should be examined in future studies. It is interesting to explore the molecular mechanisms involved in this movement. It remains uncertain whether this migration is caused by active molecules that promote hematopoietic stem cell

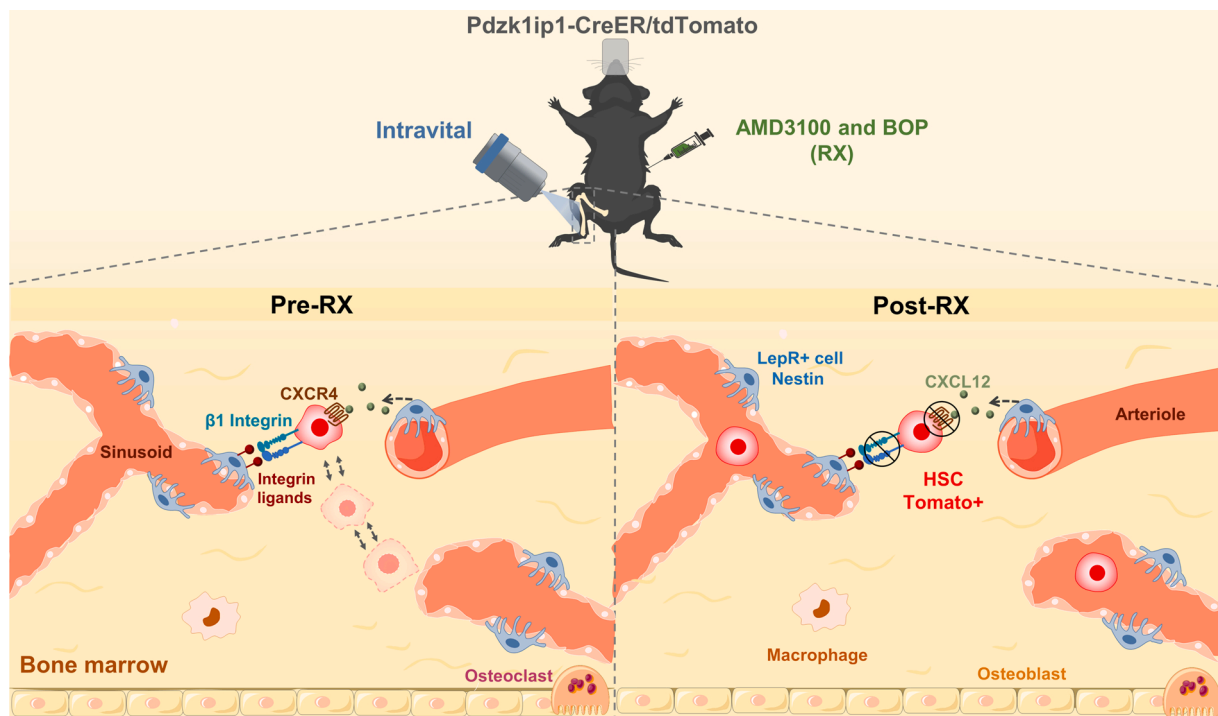


Fig. 2. Hematopoietic stem cell retained steady state within the bone marrow niche is linked to a mobile behavior.

Mobilization of the hematopoietic stem cells from the bone marrow niche by drugs, that block CXCR4 (plerixafor, AMD3100) and integrin signaling [N-(Benzenesulfonyl)-L-prolyl-L-O-(1-pyrrolidinylcarbonyl) tyrosine, (BOP)] (AMD3100 + BOP, RX), inhibits hematopoietic stem cell mobility as well as its form fluctuations within the niche (Upadhaya et al., 2020).

mobility or by the lack of specific anchoring factors. Are hematopoietic stem cells searching for a higher gradient of specific limited factors within the niche? Are other quiescent stem cells also behaving like hematopoietic stem cells in live mice? Also, as circadian rhythms influence hematopoietic stem cells (Mendez-Ferrer et al., 2009), it will be attractive to examine whether hematopoietic stem cell behavior varies during light cycles.

2.4. Interactions within the bone marrow niche

The bone marrow microenvironment defines the hematopoietic stem cell fate (Rashidi et al., 2014). Experimental data has revealed that small changes in niche regulatory mechanisms affect directly hematopoietic stem cells (Birbrair and Frenette, 2016b). Understanding exactly how hematopoietic stem cells are controlled by their niche is of fundamental importance. Upadhaya and colleagues showed the proximity of hematopoietic stem cells to the perivascular zones (Upadhaya et al., 2020), as it has been previously reported (Birbrair and Frenette, 2016b). Nevertheless, the perivascular niche itself is complex. Perivascular cells have been distinguished as essential components of the hematopoietic stem cell microenvironment (Mendez-Ferrer et al., 2010; Pinho et al., 2013), and *in vivo* genetic elimination of those cells from the bone marrow directly affects hematopoietic stem cells (Mendez-Ferrer et al., 2010). There are two main subpopulations of bone marrow perivascular cells in regards to their vascular positions: sinusoidal and arteriolar pericytes (Kunisaki et al., 2013; Nobre et al., 2021). Most of the quiescent hematopoietic stem cells reside closer to arterioles (Kunisaki et al., 2013). Upadhaya and colleagues did not determine whether their analyzes were done in the sinusoidal or arteriolar niches (Upadhaya et al., 2020). Future studies should explore whether hematopoietic stem cells behave differently in these two central niches within live mice.

Upadhaya and colleagues showed that the blockade of C-X-C motif chemokine 12 (CXCL12) signaling abrogates hematopoietic stem cell movement in the niche (Upadhaya et al., 2020). It is not clear, however,

whether this is caused by a direct or indirect effect of the drug. Is the drug acting directly on hematopoietic stem cells or on a niche component? Interestingly, sinusoidal and arteriolar niches contribute with different cytokines for the maintenance of hematopoietic stem cells. CXCL12-derived from the arteriolar niche is essential for hematopoietic stem cells, but not the one derived from the sinusoidal niche. Thus, it would be important to analyze hematopoietic stem cell behavior in response to CXCL12 deletion only from arteriolar pericytes. In contrast, SCF from the sinusoidal niche, but not from the arteriolar, seems to be essential for hematopoietic stem cell functioning. Thus, future experiments should address how distinct niche regulatory molecules affect hematopoietic stem cells' behavior in live animals.

Modern technologies provide the possibility of eliminating single cells from the tissue microenvironment and analyzing the behavior of the remaining cells (Berthiaume et al., 2018; Santos et al., 2019; Prazeres, 2020; Coimbra-Campos, 2021; Sena, 2021). Thus, it is possible to explore the effect of eliminating single components of the niche by using targeted two-photon irradiation and analyzing the effect on hematopoietic stem cells' behavior by two-photon laser scanning microscopy. Alternatively, it will be interesting to evaluate what is the effect of the death of one hematopoietic stem cell on other neighboring hematopoietic stem cells. Thus, longitudinal imaging studies may advance significantly our knowledge on hematopoietic stem cell biology in the future.

Our better understanding of hematopoietic stem cells' behavior in their normal bone marrow microenvironment leads to questions on how these cells behave in the bone marrow in different pathologies. Changes in the normal bone marrow niche may activate the appearance of pre-leukemic microenvironments (Konopleva and Jordan, 2011). How leukemic stem cells may affect this hematopoietic stem cell behavior, as well as how the leukemia stem cells themselves behave in live animals within their niches remains to be discovered.

3. Conclusion

In conclusion, the study by Upadhaya and colleagues reveals how the most illustrious residents of the bone marrow behave within their niche in live animals (Upadhaya et al., 2020). However, our understanding of the hematopoietic stem cells' behavior in their niches still remains limited, and the complexity of interactions with all niche components should be elucidated in future studies. Despite the powerful experimental transgenic models that provide proof of concept for the hematopoietic stem cell biology within the bone marrow, we are still lacking direct demonstration of hematopoietic stem cell behavior within the human bone marrow cavity. The main question for the future is whether we can translate mice research into humans. Improving the availability of human bone marrow biopsies will be essential to reach this aim. The creation of bone marrow organoids from human induced pluripotent stem cells (iPSCs) may in the future support the data provided by elegant mouse studies.

Declaration of Competing Interest

The authors indicate no potential conflicts of interest.

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References

- Acar, M., et al., 2015. Deep imaging of bone marrow shows non-dividing stem cells are mainly perisinusoidal. *Nature* 526 (7571), 126–130.
- Al-Drees, M.A., et al., 2015. Making blood: the hematopoietic niche throughout ontogeny. *Stem Cells Int.* 2015, 571893.
- Arai, F., et al., 2004. Tie2/angiopoietin-1 signaling regulates hematopoietic stem cell quiescence in the bone marrow niche. *Cell* 118 (2), 149–161.
- Asada, N., et al., 2017a. Differential cytokine contributions of perivascular hematopoietic stem cell niches. *Nat. Cell Biol.* 19 (3), 214–223.
- Asada, N., et al., 2017b. Differential cytokine contributions of perivascular hematopoietic stem cell niches. *Nat. Cell Biol.*
- Balazs, A.B., et al., 2006. Endothelial protein C receptor (CD201) explicitly identifies hematopoietic stem cells in murine bone marrow. *Blood* 107 (6), 2317–2321.
- Barminko, J., Reinholt, B., Baron, M.H., 2016. Development and differentiation of the erythroid lineage in mammals. *Dev. Comp. Immunol.* 58, 18–29.
- Baron, M.H., 2005. Early patterning of the mouse embryo: implications for hematopoietic commitment and differentiation. *Exp. Hematol.* 33 (9), 1015–1020.
- Baron, M.H., Isen, J., Fraser, S.T., 2012. The embryonic origins of erythropoiesis in mammals. *Blood* 119 (21), 4828–4837.
- Becker, A.J., Mc, C.E., Till, J.E., 1963. Cytological demonstration of the clonal nature of spleen colonies derived from transplanted mouse marrow cells. *Nature* 197, 452–454.
- Berthiaume, A.A., et al., 2018. Dynamic remodeling of pericytes in vivo maintains capillary coverage in the adult mouse brain. *Cell Rep.* 22 (1), 8–16.
- Birbrair, A., Frenette, P.S., 2016a. Niche heterogeneity in the bone marrow. *Ann. N. Y. Acad. Sci.* 1370 (1), 82–96.
- Birbrair, A., Frenette, P.S., 2016b. Niche heterogeneity in the bone marrow. *Ann. N. Y. Acad. Sci.* 1370 (1), 82–96.
- Birbrair, A., et al., 2013. Type-1 pericytes participate in fibrous tissue deposition in aged skeletal muscle. *Am. J. Physiol., Cell Physiol.* 305 (11), C1098–1113.
- Borges, I.D.T., et al., 2017. Lung as a niche for hematopoietic progenitors. *Stem Cell Reviews and Reports.*
- Bowen, J.M., et al., 2015. Extramedullary hematopoiesis in a sentinel lymph node as an early sign of chronic myelomonocytic leukemia. *Case Rep. Pathol.* 2015, 594970.
- Bowman, T.V., Zon, L.I., 2009. Lessons from the niche for generation and expansion of hematopoietic stem cells. *Drug Discov. Today Ther. Strateg.* 6 (4), 135–140.
- Bozzini, C.E., et al., 1970. Studies on medullary and extramedullary erythropoiesis in the adult mouse. *Am. J. Physiol.* 219 (3), 724–728.
- Challen, G.A., et al., 2009. Mouse hematopoietic stem cell identification and analysis. *Cytometry A* 75 (1), 14–24.
- Chen, C.-Z., et al., 2002. Identification of endoglin as a functional marker that defines long-term repopulating hematopoietic stem cells. *Proc. Natl. Acad. Sci.* 99 (24), 15468–15473.
- Cheung, T.H., Rando, T.A., 2013. Molecular regulation of stem cell quiescence. *Nat. Rev. Mol. Cell Biol.* 14 (6), 329–340.
- Coimbra-Campos, L.M.C., et al., 2021. Circulating nestin-GFP⁺ cells participate in the pathogenesis of paracoccidioides brasiliensis in the Lungs. *Stem Cell Rev. Rep.* <https://doi.org/10.1007/s12015-021-10181-3>. <https://link.springer.com/article/10.1007/s12015-021-10181-3>.
- Cooper, B., 2011. The origins of bone marrow as the seedbed of our blood: from antiquity to the time of Osler. *Proc. (Bayl Univ Med Cent)* 24 (2), 115–118.
- Dykstra, B., et al., 2006. High-resolution video monitoring of hematopoietic stem cells cultured in single-cell arrays identifies new features of self-renewal. *Proc Natl Acad Sci U S A* 103 (21), 8185–8190.
- Ema, H., et al., 2005. Quantification of self-renewal capacity in single hematopoietic stem cells from normal and Lnk-deficient mice. *Dev. Cell* 8 (6), 907–914.
- Geiger, H., de Haan, G., Florian, M.C., 2013. The ageing haematopoietic stem cell compartment. *Nat. Rev. Immunol.* 13 (5), 376–389.
- Goodell, M.A., et al., 1997. Dye efflux studies suggest that hematopoietic stem cells expressing low or undetectable levels of CD34 antigen exist in multiple species. *Nat. Med.* 3 (12), 1337–1345.
- Gray, J.V., et al., 2004. Sleeping beauty⁺: quiescence in *Saccharomyces cerevisiae*. *Microbiol. Mol. Biol. Rev.* 68 (2), 187–206.
- Inra, C.N., et al., 2015. A perisinusoidal niche for extramedullary haematopoiesis in the spleen. *Nature* 527 (7579), 466–471.
- Jacobson, L.O., et al., 1950. The role of the spleen in radiation injury and recovery. *J. Lab. Clin. Med.* 35 (5), 746–770.
- Jacobson, L.O., et al., 1951. Recovery from radiation injury. *Science* 113 (2940), 510–511.
- Johns, J.L., Christopher, M.M., 2012. Extramedullary hematopoiesis: a new look at the underlying stem cell niche, theories of development, and occurrence in animals. *Vet. Pathol.* 49 (3), 508–523.
- Khaddour, K., Hana, C.K., Mewawalla, P., 2020. Hematopoietic stem cell transplantation. StatPearls. Treasure Island (FL).
- Khan, J.A., et al., 2016. Fetal liver hematopoietic stem cell niches associate with portal vessels. *Science* 351 (6269), 176–180.
- Kiel, M.J., Radice, G.L., Morrison, S.J., 2007a. Lack of evidence that hematopoietic stem cells depend on N-cadherin-mediated adhesion to osteoblasts for their maintenance. *Cell Stem Cell* 1 (2), 204–217.
- Kiel, M.J., et al., 2007b. Hematopoietic stem cells do not asymmetrically segregate chromosomes or retain BrdU. *Nature* 449 (7159), 238–242.
- Konopleva, M.Y., Jordan, C.T., 2011. Leukemia stem cells and microenvironment: biology and therapeutic targeting. *J. Clin. Oncol.* 29 (5), 591–599.
- Kumaravelu, P., et al., 2002. Quantitative developmental anatomy of definitive hematopoietic stem cells/long-term repopulating units (HSC/RUS): role of the aorta-gonad-mesonephros (AGM) region and the yolk sac in colonisation of the mouse embryonic liver. *Development* 129 (21), 4891–4899.
- Kunisaki, Y., et al., 2013. Arteriolar niches maintain haematopoietic stem cell quiescence. *Nature* 502 (7473), 637–643.
- Laurenti, E., et al., 2008. Hematopoietic stem cell function and survival depend on c-Myc and N-Myc activity. *Cell Stem Cell* 3 (6), 611–624.
- Lefrancais, E., et al., 2017. The lung is a site of platelet biogenesis and a reservoir for hematopoietic progenitors. *Nature* 544 (7648), 105–109.
- Lewandowski, D., et al., 2010. In vivo cellular imaging pinpoints the role of reactive oxygen species in the early steps of adult hematopoietic reconstitution. *Blood* 115 (3), 443–452.
- Lewis, D.J., et al., 1994. Perirenal liposarcoma containing extramedullary hematopoiesis associated with renal cell carcinoma. *Urology* 43 (1), 106–109.
- Lo Celso, C., et al., 2009. Live-animal tracking of individual hematopoietic stem/progenitor cells in their niche. *Nature* 457 (7225), 92–96.
- Lorenz, E., et al., 1951. Modification of irradiation injury in mice and guinea pigs by bone marrow injections. *J. Natl. Cancer Inst.* 12 (1), 197–201.
- Lux, C.T., et al., 2008. All primitive and definitive hematopoietic progenitor cells emerging before E10 in the mouse embryo are products of the yolk sac. *Blood* 111 (7), 3435–3438.
- Macki, M., et al., 2013. Presacral extramedullary hematopoiesis: an alternative hypothesis. *J. Clin. Neurosci.* 20 (12), 1664–1668.
- Main, J.M., Prehn, R.T., 1955. Successful skin homografts after the administration of high dosage X radiation and homologous bone marrow. *J. Natl. Cancer Inst.* 15 (4), 1023–1029.
- Mathe, G., et al., 1959. [Transfusions and grafts of homologous bone marrow in humans after accidental high dosage irradiation]. *Rev. Fr. Etud. Clin. Biol.* 4 (3), 226–238.

- Mathe, G., et al., 1963. Haematopoietic chimera in man after allogenic (Homologous) bone-marrow transplantation. (Control of the secondary syndrome. Specific tolerance due to the chimerism). *Br. Med. J.* 2 (5373), 1633–1635.
- Medvinsky, A., Dzierzak, E., 1996. Definitive hematopoiesis is autonomously initiated by the AGM region. *Cell* 86 (6), 897–906.
- Medvinsky, A.L., et al., 1993. An early pre-liver intraembryonic source of CFU-S in the developing mouse. *Nature* 364 (6432), 64–67.
- Medvinsky, A., Rybtsov, S., Taoudi, S., 2011. Embryonic origin of the adult hematopoietic system: advances and questions. *Development* 138 (6), 1017–1031.
- Mendelson, A., Frenette, P.S., 2014. Hematopoietic stem cell niche maintenance during homeostasis and regeneration. *Nat. Med.* 20 (8), 833–846.
- Mendez-Ferrer, S., et al., 2009. Circadian rhythms influence hematopoietic stem cells. *Curr. Opin. Hematol.* 16 (4), 235–242.
- Mendez-Ferrer, S., et al., 2010. Mesenchymal and haematopoietic stem cells form a unique bone marrow niche. *Nature* 466 (7308), 829–834.
- Muller, A.M., et al., 1994. Development of hematopoietic stem cell activity in the mouse embryo. *Immunity* 1 (4), 291–301.
- Muller-Sieburg, C.E., et al., 2012. Stem cell heterogeneity: implications for aging and regenerative medicine. *Blood* 119 (17), 3900–3907.
- Nakamura-Ishizu, A., Suda, T., 2014. Aging of the hematopoietic stem cells niche. *Int. J. Hematol.* 100 (4), 317–325.
- Nobre, A.R., Risson, E., Singh, D.K., et al., 2021. Bone marrow NG2⁺/Nestin⁺ mesenchymal stem cells drive DTC dormancy via TGF- β 2. *Nat. Cancer* 2, 327–339. <https://doi.org/10.1038/s43018-021-00179-8>.
- Okada, S., et al., 1992. In vivo and in vitro stem cell function of c-kit- and Sca-1-positive murine hematopoietic cells. *Blood* 80 (12), 3044–3050.
- Palis, J., et al., 1999. Development of erythroid and myeloid progenitors in the yolk sac and embryo proper of the mouse. *Development* 126 (22), 5073–5084.
- Pang, W.W., et al., 2011. Human bone marrow hematopoietic stem cells are increased in frequency and myeloid-biased with age. *Proc. Natl. Acad. Sci. U. S. A.* 108 (50), 20012–20017.
- Pearce, D.J., et al., 2004. Multiparameter analysis of murine bone marrow side population cells. *Blood* 103 (7), 2541–2546.
- Pinho, S., et al., 2013. PDGFR α and CD51 mark human nestin⁺ sphere-forming mesenchymal stem cells capable of hematopoietic progenitor cell expansion. *J. Exp. Med.* 210 (7), 1351–1367.
- Prazeres, P., et al., 2020. Ablation of sensory nerves favours melanoma progression. *J. Cell. Mol. Med.* 24 (17), 9574–9589. <https://doi.org/10.1111/jcmm.15381>. <https://onlinelibrary.wiley.com/doi/full/10.1111/jcmm.15381>.
- Rashidi, N.M., et al., 2014. In vivo time-lapse imaging shows diverse niche engagement by quiescent and naturally activated hematopoietic stem cells. *Blood* 124 (1), 79–83.
- Rhodes, K.E., et al., 2008. The emergence of hematopoietic stem cells is initiated in the placental vasculature in the absence of circulation. *Cell Stem Cell* 2 (3), 252–263.
- Sanchez-Aguilera, A., Mendez-Ferrer, S., 2017. The hematopoietic stem-cell niche in health and leukemia. *Cell. Mol. Life Sci.* 74 (4), 579–590.
- Santos, G.S.P., et al., 2019. Pericyte plasticity in the brain. *Neurosci. Bull.* 35 (3), 551–560.
- Schnuelle, P., et al., 1999. Idiopathic myelofibrosis with extramedullary hematopoiesis in the kidneys. *Clin. Nephrol.* 52 (4), 256–262.
- Schofield, R., 1978. The relationship between the spleen colony-forming cell and the haemopoietic stem cell. *Blood Cells* 4 (1–2), 7–25.
- Sena, I.F.G., et al., 2021. C(3)1-TAG in C57BL/6 J background as a model to study mammary tumor development. *Histochem. Cell Biol.* <https://doi.org/10.1007/s00418-021-01995-w>. <https://link.springer.com/article/10.1007/s00418-021-01995-w>.
- Siminovitch, L., McCulloch, E.A., Till, J.E., 1963. The distribution of colony-forming cells among spleen colonies. *J. Cell. Comp. Physiol.* 62, 327–336.
- Singh, A.K., Cancelas, J.A., 2020. Gap junctions in the bone marrow lymphohematopoietic stem cell niche, leukemia progression, and chemoresistance. *Int. J. Mol. Sci.* 21 (3).
- Sohawon, D., et al., 2012. Extra-medullary haematopoiesis: a pictorial review of its typical and atypical locations. *J. Med. Imaging Radiat. Oncol.* 56 (5), 538–544.
- Sugiyama, D., Tsuji, K., 2006. Definitive hematopoiesis from endothelial cells in the mouse embryo; a simple guide. *Trends Cardiovasc. Med.* 16 (2), 45–49.
- Swain, A., et al., 2014. Intrinsic and extrinsic regulation of mammalian hematopoiesis in the fetal liver. *Histol. Histopathol.* 29 (9), 1077–1082.
- Takizawa, H., et al., 2011. Dynamic variation in cycling of hematopoietic stem cells in steady state and inflammation. *J. Exp. Med.* 208 (2), 273–284.
- Tanaka, Y., et al., 2015. Embryonic hematopoietic progenitor cells reside in muscle before bone marrow hematopoiesis. *PLoS One* 10 (9), e0138621.
- Tavian, M., Peault, B., 2005. Embryonic development of the human hematopoietic system. *Int. J. Dev. Biol.* 49 (2–3), 243–250.
- Thomas, E.D., et al., 1957. Intravenous infusion of bone marrow in patients receiving radiation and chemotherapy. *N. Engl. J. Med.* 257 (11), 491–496.
- Till, J.E., Mc, C.E., 1961. A direct measurement of the radiation sensitivity of normal mouse bone marrow cells. *Radiat. Res.* 14, 213–222.
- Tsamandas, A.C., et al., 1995. Extramedullary hematopoiesis in the allograft liver. *Mod. Pathol.* 8 (6), 671–674.
- Upadhaya, S., et al., 2020. Intravital imaging reveals motility of adult hematopoietic stem cells in the bone marrow niche. *Cell Stem Cell*.
- Vassiliou, V., et al., 2012. Presacral extramedullary hematopoiesis in a patient with rectal adenocarcinoma: report of a case and literature review. *J. Gastrointest. Cancer* 43 (Suppl 1), S131–135.
- Wayne, A.S., Baird, K., Egeler, R.M., 2010. Hematopoietic stem cell transplantation for leukemia. *Pediatr. Clin. North Am.* 57 (1), 1–25.
- Wolf, N.S., Trentin, J.J., 1970. Differential proliferation of erythroid and granuloid spleen colonies following sublethal irradiation of the bone marrow donor. *J. Cell. Physiol.* 75 (2), 225–229.
- Woodward, N., et al., 2000. Renal myelofibrosis: an unusual cause of renal impairment. *Nephrol. Dial. Transplant.* 15 (2), 257–258.
- Wu, A.M., et al., 1967. A cytological study of the capacity for differentiation of normal hemopoietic colony-forming cells. *J. Cell. Physiol.* 69 (2), 177–184.
- Wu, A.M., et al., 1968. Cytological evidence for a relationship between normal hemopoietic colony-forming cells and cells of the lymphoid system. *J. Exp. Med.* 127 (3), 455–464.
- Yamamoto, S., et al., 2020. Hematopoietic stem cell transplantation for pediatric acute promyelocytic leukemia in Japan. *Pediatr. Blood Cancer* 67 (5), e28181.
- Yang, L., et al., 2005. Identification of Lin(-)Sca1(+)-kit(+)-CD34(+)-Flt3- short-term hematopoietic stem cells capable of rapidly reconstituting and rescuing myeloablated transplant recipients. *Blood* 105 (7), 2717–2723.
- Yoder, M.C., 2004. Blood cell progenitors: insights into the properties of stem cells. *Anat. Rec. A. Discov. Mol. Cell. Evol. Biol.* 276 (1), 66–74.

Walison N. Silva is a Master student at Department of Pathology at the Federal University of Minas Gerais, Brazil, pursuing research in melanoma microenvironment and stem cells.

Alinne C. Costa, M.Sc., is a Ph.D. student in the Department of Pathology at the Federal University of Minas Gerais. Her research focuses on skeletal muscle regeneration, muscle development, stem cells, and cancer biology.

Caroline C. Picoli, M.Sc., is a Ph.D. student at the Department of Pathology at the Federal University of Minas Gerais. Her research focuses on the microenvironment of adipose perivascular cells.

Beatriz G. S. Rocha, is a Master student at Department of Pathology at the Federal University of Minas Gerais, pursuing research in perivascular cells influence on breast cancer progression.

Gabryella S. P. Santos, M.Sc. is a Ph.D. student in the Department of Pathology at the Federal University of Minas Gerais. Her research focuses on research in the prostate tumor microenvironment.

Pedro A. C. Costa, M.Sc., Ph.D., is a postdoctoral researcher in the Department of Pathology at the Federal University of Minas Gerais. He obtained his Ph.D. in Health Sciences from the René Rachou Research Center (CPqRR-Fiocruz Minas). His research focuses on the immunoregulation of the tumor microenvironment.

Parviz Azimnasab-sorkhabi M.Sc., is pursuing a Ph.D. in the Pathology Department at the Federal University of Minas Gerais. His research focuses on the role of cancer stem cells in the tumor microenvironment.

Maryam Soltani-asl, M.Sc., is a Ph.D. student at the Department of Pathology at the Federal University of Minas Gerais. Her research focuses on the melanoma metastasis microenvironment.

Rodrigo A. da Silva, M.Sc., Ph.D., is a postdoctoral researcher in the Department of Pathology at the Federal University of Minas Gerais. He obtained his Ph.D. in Biochemistry from Campinas State University. His research focuses on biochemistry, molecular biology, and mechanisms of gene regulation mediated by epigenetic mechanisms.

Jaime Henrique Amorim M.Sc., Ph.D., is a professor in the Department of Biochemistry and Immunology at the Federal University of West Bahia. He obtained his Ph.D. in Biotechnology at the University of São Paulo. His main research interests focus on vaccine development for cancer.

Rodrigo R. Resende, M.Sc., Ph.D., is a professor in the Department of Biochemistry and Immunology at the Federal University of Minas Gerais. He obtained his Ph.D. in Biochemistry from the University of São Paulo. His research focuses on Neuroscience, Nanobiotechnology, Stem Cells, Calcium Signaling, and Tissue Engineering.

Akiva Mintz, M.D., Ph.D., is a Professor of Radiology at Columbia University Medical Center and Attending Radiologist at New York Presbyterian Hospital. He obtained his medical and graduate degrees at the Pennsylvania State University College of Medicine. Dr. Mintz's cross-translational research efforts exploit nuclear-based molecular imaging and therapy techniques to personalize anti-cancer therapies.

Alexander Birbrair, Ph.D., is a Professor in the Department of Pathology at the Federal University of Minas Gerais. He obtained his Ph.D. in Neuroscience from Wake Forest School of Medicine. His laboratory is interested in understanding how the cellular components of different tissues function and control disease progression. His research is funded by the Serrapilheira Institute, CNPq, CAPES, and FAPESP. In 2018, Alexander was elected affiliate member of the Brazilian Academy of Sciences (ABC), and, in 2019, he was elected member of the Global Young Academy (GYA).